

July, 2021

Comparative transcriptomic analysis provides new insights into graft compatibility/incompatibility in *Citrus*

Wen He^{1,2}, Rui Xie^{1,2}, Yan Wang^{1,2}, Qing Chen¹, Hao Wang^{1,2}, Shaofeng Yang^{1,2}, Jing Zhang^{1,2}, Ya Luo¹, Yong Zhang¹, Haoru Tang^{1,2}, Frederick G. Gmitter Jr³, Xiaorong Wang^{1,2,*}

¹College of Horticulture, Sichuan Agricultural University, Chengdu 611130, Sichuan, P.R. China,

²Institute of Pomology and Olericulture, Sichuan Agricultural University, Chengdu 611130, Sichuan, P.R. China

³ Citrus Research and Education Center, University of Florida, Lake Alfred 33850, Florida, USA

ABSTRACT

Grafting is a useful cultivation technology to resist abiotic and biotic stresses and is an integral part of citrus production. However, some widespread utilized rootstocks may still cause graft incompatibility in the orchard. *Citrus grandis* (L.) Osbeck cv. Hongmian miyou is mutated from 'Guanxi miyou', but these two scions showed different compatible with available Poncirus trifoliata rootstock. In the present study, the morphological and cytological development before and during foliage etiolation was first investigated, and the crucial developmental stages were defined. Afterward, with combined physiological and biochemical data and transcriptome profiling, two rate-limiting genes, NCED3 (9-cisepoxycarotenoid dioxygenases 3) and NCED5, responsible for abscisic acid accumulation were highlighted. Later, correlation analysis between coexpression modules and traits indicated that abscisic acid is the most likely inducer of the expression of stresses-related genes. In addition, the excessive starch accumulation was observed in leaf lamina and midribs of graft incompatible plant leaves. Taken together, our work provides a new insight into the role of the carotenoid and abscisic acid biosynthesis genes in regulation and contributing to the graft incompatibility, and will further efforts to define and deploy useful genes to study the mechanisms underlying citrus rootstock- scion interactions.







Fig. 2 Genes differentially expressed analyses. a Venn diagrams of genes differentially expressed between the Hm/Pt and controls at the same developmental stage. b KEGG enrichment analysis of the DEGs. c expression of DEGs in carotenoid biosynthesis and plant hormone signal transduction pathways. Heatmap color indicates FPKM value.



Fig. 3 WGCNA of differentially expressed genes. a Hierarchical cluster tree showing co-expression modules identified by WGCNA. Each leaf in the tree is one gene. The major tree branches constitute 7 modules labelled by different colors. b KEGG enrichment analysis of the genes in the module blue. c genes whose expression is highly correlated in the module blue.

CONCLUSION

In this study, we compared the transcriptomes of the leaves of graft incompatible/ compatible combinations. The results indicated that a few DEGs may cause graft incompatible happen. The current study revealed the physiological and molecular mechanisms underlying the etiolation process in citrus, and will help elucidate the mechanisms of graft incompatibility in citrus.

Fig. 4 Transverse section showing changes of leaf of different graft combinations. a Leaf samples at P3. b Cross section of leaf. c Midrib section was observed and photographs taken under light microscope. d Epifluorescence photomicrographs of phloem. e Starch grains were dyed blue. f-g The content of starch and soluble sugars. h Transcript abundance changes of starch and sucrose metabolism pathways. Asterisks represent remarkable differences compared to the control (p < 0.05, p < 0.01), analyzed using Student's t-test. Heatmap shows the log10 (FPKM+0.01) of selected differentially expressed transcripts. The black arrows indicate the parenchyma cells and red arrows indicate starch accumulation.

ACKNOWLEDGEMENTS

This work was financially supported by Sichuan Science and Technology Program (2019YFH0061, 2020ZHCG0027, 2019NZZJ0015).